

Blood Samples for Rhino Survival Research Project

Veterinarians asked to attend to any injured rhino are requested to first contact the primary researcher (Jacques du Preez - 072 402 3126) to try and arrange for him to go with you to the rhino to collect his samples. If not, follow the procedure below:

Step 1

Stabilise patient

Samples needed:

Step 2

Collect blood from the auricular vein in 4 ml vacuum tubes using vacutainer needle and hub. Tubes must be as full as possible please.

- 2 x Serum tubes (red)
- 1 x EDTA tube (purple)
- 2 x Heparin tubes (green)
- 1 x Fluoride oxalate tube (grey)
- 1 x Citrate tube (light blue)
- 2 x thin peripheral blood smears, unfixed and unstained

Do NOT put on ice directly at this stage, but keep it cool.

Samples must be kept out of direct sunlight, upright and with as little physical disturbance (shaking etc.) as possible.

Step 3

Take Lactate measurements every 20 minutes

Machine available from Dr Steenkamp free of charge for use in this project only.

Procedure of how to use the machine will be demonstrated by Jacques du Preez to anyone willing to be involved with this project.

Only use verified handheld machine

All serial lactate measurements must be carefully recorded as demonstrated and results sent to the OVAH Clinical Pathology lab together with a full clinical history, contact details of persons involved, patient ID and the blood samples as prescribed.

Step 4

Processing blood samples:

1. Centrifuge Serum and Heparin tubes at 4000rpm for 8 min.
2. Remove serum and plasma from the cells.
3. Freeze (preferred) or refrigerate serum and plasma.
4. 2 thin blood smears of EDTA blood, unstained and unfixed, properly labeled.

Step 5

Send everything to the lab:

Attention: Prof Amelia Goddard
Clinical Pathology Laboratory
Onderstepoort Veterinary Academic
Hospital, R3-11
Old Soutpan Road
Onderstepoort, Pretoria
0110